

GENE NETWORK INVOLVED IN SEX DETERMINATION- A REVIEW

¹Pandey C, ²Shukla PR, ³Vishvkarma R, ⁴Bisen JS

¹R &D Division, MRD LifeSciences Pvt. Ltd., Lucknow, Uttar Pradesh, India.

²Division of Plant Biotechnology, Central Institute of Medicinal and Aromatic Plants,
Lucknow, India

³Division of Endocrinology, Central Drug Research Institute, Lucknow, India

⁴Shri Rama Krishna College of Commerce and Science, Satna, Madhya Pradesh, India

***Corresponding Author: ChitranshuPandey**

Email ID: chitranshupandeyniu@gmail.com

Available online at: www.ijbbas.in.

Received 20th March. 2020; Revised 24th March. 2020; Accepted 5th April. 2020; Available online: April. 2020

ABSTRACT

Sex developmental disorders (SDDs) are inborn conditions where the changes of gonadal, or physiological sex is anomalous. A majority of the genes necessary for genital evolution were characterized through the investigation of disorder of sex determination mice. Nevertheless, the use of knockout and mutated mouse strains has greatly helped to explore the role and relationships of gonad genes within the network of the developmental genes. However, in recent years, the genetic help of defining and differentiating mammalian sex has progressed a lot. Here we study our current understanding of determining mice's sex based on observations from mouse models.

Key word: Chromosomes, LIM homeobox, Sex developmental disorders, Gonad genes.

INTRODUCTION

Genetic differences among sexes during embryo development are genetically resolute among the primates. These disparities affect human and physically physiological, sexual and social life significantly. Genetic differences among sexes throughout embryo development are genetically resolute among the primates. These disparities have massive psychological, cultural, and social effects on human and physical life. Sex creation can be categorized into two types of acts,' sex determination,' which is embryonic determination that governs the indistinct gonads that behave either ovaries or testicles, and' sex determination,' which occurs as the gonad develops and is determined to decide the phenotypical sex by the gonad outcome. Sex resolution, in primates, is equivalent to genital development. The factors that affect sex identification tend which regulate transcription, in which secreted hormones and their receptors are referred as factors affecting sex variations. Most of our knowledge of genes and their related proteins in sex-making comes from mouse models. There the raging understanding of gonad formation would be

discussed, and how alteration of this dynamic growth network leads to the disorder of sex development. Gens that have participated in the development of embryonic mouse gonad need to be given special consideration.

Gonadal development in mice

Among rodents, both female and male are complemented by 19 sets of autosomal chromosomes but not similar among their sex chromosomes. Men contain one X and another Y (40, XY) among rodents, while females have two X chromosomes (40, XX). Two-sex embryos differ only by sex chromosomes at the time of conception. The first proof of dimorphism of sex in rodent embryos is significant at the time of the formation of bi-potential gonad to grow into either an ovary or a testis in XX and XY people. The commitment is produced in mice around embryonic day 10.5. Gonad distinction helps in ovarian and testicular hormone secretion and psychological activation. SRY's function in XY gonads is to tip the scale toward the route, which is unique to the testis [1, 2].

In mice, the SRY gene is located in the hetero-potential XY gonad [3], and the SRY gene expression starts regulating SOX9 development. Sox9 has been expressed in mice to promote the production of Fgf9 and then both sox9 and FGF9 function together in +ve feedback loops, that are often postulated to repress wnt4 and are geared towards setting up testis-specific pathways. In the non-appearance of SRY in XX species, WNT 4 and RSPO1 are produced at high range and control cytoplasmic β -catenin, which is then moved into the nucleus, where it binds to the transcription factor / lymphoid enhancer-binding factor and turns on the transcription of the target gene. Both WNT4 and beta-catenin inhibit SOX9/FGF9's positive feedback loop, allowing for specific ovarian pathways to be formed.

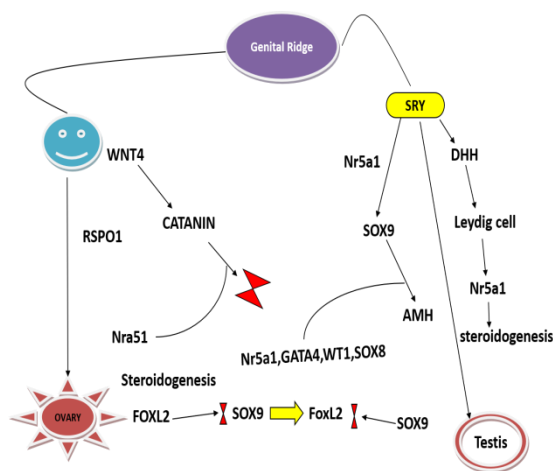


Fig1. Genomic cascade in sex determination.

Gene compulsory for establishment of Bi-potential gonad-

Mammalian gonads from bi-potential gonad or vaginal ridges are visible in both sexes [4, 5, 6]. In mouse, they first appear in specific region of mesonephros at around embryonic days E9.5, one day before the beginning of SRY speech, which is necessary to facilitate the formation of testis in XY individuals. Diverse genes have shown their significance in bi-potential gonad growth. The development of genital ridges takes place via coelomic epithelium and afterwards via split into fragments of underlie basement membrane, allowing separation into layers of proliferating coelomic epithelium into inner mesenchymal region of mesonephros [7]. The rigidity of urogenital ridges starts at E10.3, then at the anteroposterior axis. Urogenital ridges are filled by moving precursors of gamete cells, combined with stiffness [8,9, 10, 11,12]. The study of mutant mice has recognized numerous significant factors in both cell proliferation and consumption. To our information, GAT4 is preliminary factor of transcription expressed mainly in urogenital ridges.

GATA4 exists in coelomic epithelium cells of rostral part of genital ridges as soon as E10.0 and expression of it spread across rostro caudal axis till embryonic 10.2 [7]. There are following genes are involved in genital ridge formation.

Table 1. Summarization of some gene's importance for testis and ovary development

| Testis | | |
|---------------|-------------------------|----------------------|
| Genes | Gene product | Mutant type |
| WT1 | Factor in Transcription | Gonadal agenesis |
| SF1 | Nuclear receptor | GR development block |
| GATA 4 | Factor in Transcription | XY Reversal of Sex |
| SRY | Factor in Transcription | XY Reversal of Sex |
| Sox9 | Factor in Transcription | XY Reversal of Sex |

| Fgf9 | Signaling molecule | XY Reversal of Sex |
|----------------|-------------------------|------------------------------------|
| Dax1 | Nuclear receptor | XY hypogonadism |
| DHH | Signaling molecule | XY Gonadal dysgenesis |
| DMRT1 | Factor in Transcription | XY female |
| AMH | Peptide hormone | Persistent Mullerian duct syndrome |
| Ovaries | | |
| WNT4 | Signaling molecule | Mullerian duct agenesis |
| FOXL2 | Factor in Transcription | Premature ovarian failure |
| DAX1 | Nuclear receptor | XY sex reversal |

Nr5a1

super family of nuclear receptor adrenal 4 linking protein is also known as nuclear receptor subfamily 5, group A, category 1(Nr5a1), and transcription factor 1(Sf1) involved in sex determination was first defined as a gene coding a general transactivating factor linked to the steroid hydroxylase gene promoting it. Nr5a1+ve progenitor cells originate in adrenal glands and the coelomic epithelium somatic branch of gonads. Our adrenal glands are not well formed and exhibit fewer cell proliferation in mouse embryos [13]. Compound heterozygote study of six 1+/-, six 4+/- and Nr5a1+/-embryos reveals the disrupted development of progenitor gonadal cells depending on the expression level of Nr5a1 [14]. Embryos are those with homozygous deletion of Nr5a1 express gonad regression by embryonic 12.5 with genetically programmed cell death of gonadal somatic cells [15, 16, 17]. Conversely, an enhanced amount of fetal Leydig cells [18] accomplishes an irregular production of Nr5a1 in Pod 1 lacZ / lac Z gonads. Therefore, over expression of Nr5a1 in Nr5a1 negative mice save compromised development of gonad and spleen. This difference in the rescue effect could convey partly the differential degree of expression of Nr5a1 between tissues and the

distinctive responsiveness to the dosage of genes. [19]. Nr5a1 works in gene cluster activation participate in steroidogenesis, such as cyp 17a1 and 3beta-HSD in testicular Leydig cells. However, Nr5a1 was first determined as a gene encoding a steroidogenic gene, agent that increase rate of gene expression [20, 21, 22]. There are also numerous physiological processes in Nr5a1 [23, 24]. It could be proposed that Nr5a1 regulates the transcription of reference gene clusters involved in various physiological acts, including cell proliferation stimulation, metabolism, segregation and persistence that are critical for gonadal growth. The development of damaged gonadal and glands of adrenal in insulin / IGF signaling pathways in embryos is favored for this postulate. Insulin / IGF signaling mechanism is found to regulate the various physiological activities [19]. At a late stage, Nr5a1 controls the transcription of these important genes in XY gonad development for testicular differentiation, such as SRY-associated HMG box 9(Sox9) and anti-Mullerian hormone [25, 26]. A subset of larger population of coelomic epithelial cells expressing Nr5a1 and Gata4 is called the earlier population that belongs to genital ridge somatic lineages.

Such findings indicate that the creation of an exceptionally long narrow genital ridge concludes with Gata4 completing the anterior portion of the coelomic epithelium and Nr5a1 expression.

SRY and Sox9

The fate of embryonic gonad is set on by dividing somatic cells into Sertoli cells or granulose cells. The proliferation of Sertoli cells in bi-potential gonads enhances the testicular differentiation strategy by enhancing the ovarian differentiation system. The function of the gene part Y linked gene SRY displaces the bipotential embryonic gonad to a testicular destiny to the greatest extent of the determination of mouse sex [27, 28, 1]. The immediate role of SRY is to cause pre-Sertoli differentiation of cells, which is necessary for the dedifferentiation of bipotential gonads from the testis. Spatiotemporal expression regulated in the Sertoli cells precursors SRY. In mouse genital ridges, SRY if initially expressed on E11.0, achieves high degree of expression at E11.5 and is lost shortly after E12.5. SRY's speech starts in the center of the genital ridges and becomes restricted to the posterior area before disappearing completely in the genital ridges. Sox9 is expressed in Sertoli cell

precursors approximately 4 hours after SRY expression starts [29, 30, 31, 32, 33]. To activate the male development plan, the output of either sox9 or SRY in bipotent gonad has been studied for transgenic mouse [28, 34, 35, 36]. Sox9 and SRY are transcription factors with a known amino acid sequence as the domain HMG [37, 38]. This HMG motif appears to grab DNA sequences (A / T) ACAA(T / A) with high avidity of sox family proteins [39]. In the highest degree SRY deficient find human patients showing male to female sex transfer that affects SRY's ability to bind and bend DNA [39, 40]. SRY's deficient analysis of its C terminal domain suggests that SRY C terminal domain could contribute to SRY conformation and could affect SRY functions [41]. Nuclear localization signal at the north end of the SRY HMG box and SRY disabled in this NLS contributes to the elimination of nuclear imports, which explains only a few cases of human sex reversal [42, 43].

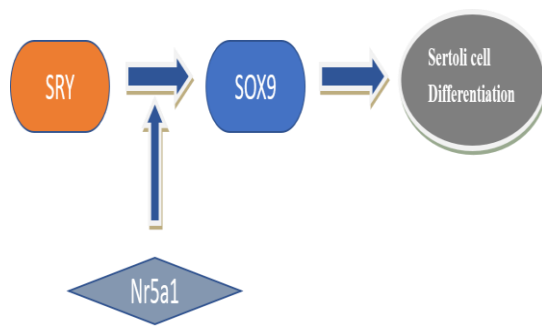


Fig 2. SRY and the transcriptional network that controls testis determination. In mice, having one specific genetic source, the Y-linked gene SRY, induces differentiation of pre-Sertoli cells. SRY directly transactivates Sox9 via the core element of the Sox9 (Tesco) testis-specific enhancer region, together with Nr5a1. Sox9 itself, along with Nr5a1, also leads to Tesco's protection of the Sox9 expression.

Instead of SRY, Sox9 transgene was accomplished to improve the technique of the method of testicular differentiation [44, 45]. Hence the important action of SRY in deciding the testis can only be over Sox9 voice. Another risk is that the Sox9 transgene drug promotes not only endogenous Sox9, but also other SRY target genes that are required to facilitate the test growth.

WT1

Wilms' tumor 1 (WT1) is a gene that encodes

the transcriptional factor of the zinc finger, was first noticed as the Wilms tumor responsible gene. Especially in children, kidney cancer is caused by human WT1 mutation [46]. WT1 contains two isoforms with additional three amino acids (threonine, lysine, serine) between the third and fourth zinc digits. WT1 is mostly found in mesodermal developmental tissues such as the urogenital slope, gonads Armstrong et al. 1993. The gonadal phenotype is replicated in other disorders where improvements in WT1 such as Denys-Drash syndrome (gonadal defects and renal failure) have been made [47, 48] and Frasier syndrome [49]. The Frasier sufferer was shown to undergo modifications leading to loss of isoform WT1 + KTS. In these isoforms, these three specific amino acids exist between the third and fourth zinc fingers of WT1, lysine, threonine, and serine. The 2 distinct isoforms –kts and +kts were expressed in the embryogenesis process in order to play different functions. It was shown that the –KTS isoforms fastened to the SRY promoter region which led to SRY transactivation [50]. In fact, the +kts WT1 isoforms were manifested within the promoter Nr5a1 to fasten sequences [51].

In addition, isoforms with + kts also play an important role in regulating SRY transcript. Removal of this+ kts of WT1 isoforms leads to reduced SRY degree in all probability driven by binding affinity to RNA. In fact, the-ktsWT1 isoforms were manifested within the promoter Nr5a1 to fasten sequences [51]. In addition, isoforms with + kts also play an important role in regulating SRY transcript. Removal of this+ kts of WT1 isoforms leads to reduced SRY degree in all probability driven by binding affinity to RNA [52].

Emx2

The gene, analogous to empty spirals from the *Drosophila* head gap gene, is Empty Spiracles homeobox 2. Lacking Emx2 in mouse embryos leads to deformation or agenesis of ureters, kidneys, and gonads, and genital tract, while normal bladder and adrenal gland development. [53] Coelomic epithelium cells are lowering their cell polarity by forming Emx2-/genital ridges. The number of migrated Emx2-/coelomic epithelial cells in the mesenchymal compartment also decreases, leading to the formation of damaged gonads [54].

LIM Homeobox 9

The LIM homeobox 9 (LHX9) is a member of the LIM homeobox gene family. The germ cells usually move in mice that lack Lhx9 gene but the genital ridge somatic cell was unsuccessful in being able to proliferate and produce separate gonad. Typically, male genetically engineered mice, lack of antimuller hormone and testosterone, develop as phenotypic females [15, 55]. In the Lhx9 negative genital ridge, the effect of steroidogenic factor 1, a nuclear receptor necessary for gonadogenesis, is reduced to a minimum level, indicating that Lhx9 may remain upstream of Nr5a1 in an evolving mouse cascade [55]. Analysis therefore showed that LHX9 would fasten and transactivate the promoter Nr5a1 in competition with WT1 [51]. Nevertheless, LHX9 mutation review did not reveal any mutations in a number of patients with human sex determination disorder [56].

Desert Hedgehog

Desert hedgehog belongs to the class of signalling hedgehog molecules which include sonic hedgehog and Indian hedgehog [57].

The Desert hedgehog is unique among the three members of the family, represented in the development of the XY mouse gonad somatic cell community from embryo 11.5(E11.5) and continuing to sertoli cells. Even no expression is shown in XX ovaries at either time [58, 59, 60]. Desert hedgehog fasten to its patched 1 receptor (PTCH1), soon after DHH. Patched 1 receptor is linked to Leydig membrane and peritubularmyoid cells, and its level of expression increased with DHH [61,62].DHH appears to be essential in Leydig cells for over-expression of sf1 [62]. Some DHH modifications have been rendered in patients with full or partial gonadal dysgenesis levels 40, XY.

Fibroblast growth factor 9

FGF9 codes the cell survival, cell proliferation, cell differentiation, cell migration, fibroblast growth factor 9 that plays a role in various processes of evolution. Fgf9 is regulated in the bi-potential gonad, just after the presentation of SRY. Mice lacking in Fgf9 demonstrate reversal of male to female sex along with impaired cell growth of the sertoli. Although this is evident on some inherited cases but not on another [63, 64]. Because of the lack of Fgf9, sox9 activity is not controlled,

Sertoli cells do not distinguish, testis development is stopped and somatic cell genes instead express ovarian growth and female pathway. There is a research in support of the idea that SRY and Nr5a1 start positive feedback loop by over-expression sox9, that is, turn up regulates FGF9 and further enhances the expression of sox9 [65]. FGFR2 is integrated in progenitor Sertoli cells plasma membrane and interprets the developmental check for Sertoli cell proliferation and differentiation [66].

CONCLUSION

After the discovery of SRY in 1990, our understanding of genes and its expression was indulging in genitalprogress and Sex-determination causative agents has been greatly enhanced. Comparative genomic hybridization (CGH)/SNP chip arrays continue to analyze ideal gene rearrangements and engage in differentiating and evaluating gonads with the development of better protocols.However, even mysterious DSD situations, more split in the testis and ovary production network shows us that there is still much to perceive.

Such 2 array-based methods require rearrangements to be assessed in presumed regulatory elements that, if changed, would contribute to Disorder of Sex Determination. The usage of microarrays to test Copy Number Variation has had some favorable results in the identification of causative mutations in mysterious DSD situations. This method, though, is restricted to determining significant rearrangements and may fail to notice small genomic rearrangements or point mutations. Next-generation sequencing (NGS) and whole-genome sequencing can have a higher purpose for mutation identification and should help to overcome mysterious DSD events. Whole genome sequencing, which encompasses all other methods, is an important tool for evaluating.

REFERENCES

- [1] Sinclair, A. H., Berta, P., Palmer, M. S., Hawkins, J. R., Griffiths, B. L., Smith, M. J., ... & Goodfellow, P. N. (1990). A gene from the human sex-determining region encodes a protein with homology to a conserved DNA-binding motif. *Nature*, 346(6281), 240-244.
- [2] Lovell-Badge, R. (1992). The role of Sry in mammalian sex determination. In *Ciba Found. Symp* (Vol. 165, pp. 162-79).
- [3] Koopman, P., & Munsterberg, A. C. B. Vivian, N. and Lovell-Badge. R. 1990. *Expression of a candidate sex determining gene during mouse testis differentiation. Nature*, 348, 150-152.
- [4] Capel, B. (1998). Sex in the 90s: SRY and the switch to the male pathway. *Annual review of physiology*, 60(1), 497-523.
- [5] Swain, A., & Lovell-Badge, R. (1999). Mammalian sex determination: a molecular drama. *Genes & development*, 13(7), 755-767.
- [6] Capel, B. (2000). The battle of the sexes. *Mechanisms of development*, 92(1), 89-103.
- [7] Hu, Y. C., Okumura, L. M., & Page, D. C. (2013). Gata4 is required for formation of the genital ridge in mice. *PLoS genetics*, 9(7).
- [8] Richardson, B. E., & Lehmann, R. (2010). Mechanisms guiding primordial germ cell migration: strategies from different organisms. *Nature reviews Molecular cell biology*, 11(1), 37-49.
- [9] Molyneaux, K., & Wylie, C. (2004). Primordial germ cell migration. *International Journal of Developmental Biology*, 48(5-6), 537-543.

- [10] Raz, E. (2004). Guidance of primordial germ cell migration. *Current opinion in cell biology*, 16(2), 169-173
- [11] Wylie, C. C., Stott, D., & Donovan, P. J. (1986). Primordial germ cell migration. In *The Cellular Basis of Morphogenesis* (pp. 433-448). Springer, Boston, MA.
- [12] Wylie, C. (1999). Germ cells. *Cell*, 96(2), 165-174.
- [13] Bland, M. L., Fowkes, R. C., & Ingraham, H. A. (2004). Differential requirement for steroidogenic factor-1 gene dosage in adrenal development versus endocrine function. *Molecular Endocrinology*, 18(4), 941-952.
- [14] Fujimoto, Y., Tanaka, S. S., Yamaguchi, Y. L., Kobayashi, H., Kuroki, S., Tachibana, M., ...&Nishinakamura, R. (2013). Homeoproteins Six1 and Six4 regulate male sex determination and mouse gonadal development. *Developmental cell*, 26(4), 416-430.
- [15] Luo, X., Ikeda, Y., & Parker, K. L. (1994). A cell-specific nuclear receptor is essential for adrenal and gonadal development and sexual differentiation. *Cell*, 77(4), 481-490.
- [16] Sadovsky, Y., Crawford, P. A., Woodson, K. G., Polish, J. A., Clements, M. A., Tourtellotte, L. M., ... &Milbrandt, J. (1995). Mice deficient in the orphan receptor steroidogenic factor 1 lack adrenal glands and gonads but express P450 side-chain-cleavage enzyme in the placenta and have normal embryonic serum levels of corticosteroids. *Proceedings of the national academy of sciences*, 92(24), 10939-10943.
- [17] Shinoda, K., Lei, H., Yoshii, H., Nomura, M., Nagano, M., Shiba, H., ...&Morohashi, K. I. (1995). Developmental defects of the ventromedial hypothalamic nucleus and pituitary gonadotroph in the Ftz-F1 disrupted mice. *Developmental Dynamics*, 204(1), 22-29.
- [18] Cui, S., Ross, A., Stallings, N., Parker, K. L., Capel, B., &Quaggin, S. E. (2004). Disrupted gonadogenesis and male-to-female sex reversal in Pod1 knockout mice. *Development*, 131(16), 4095-4105.
- [19] Efstratiadis, A. R. G. I. R. I. S. (2004). Genetics of mouse growth. *International Journal of Developmental Biology*, 42(7), 955-976.

- [20] Honda, S. I., Morohashi, K. I., Nomura, M., Takeya, H., Kitajima, M., & Omura, T. (1993). Ad4BP regulating steroidogenic P-450 gene is a member of steroid hormone receptor superfamily. *Journal of Biological Chemistry*, 268(10), 7494-7502.
- [21] Lala, D. S., Rice, D. A., & Parker, K. L. (1992). Steroidogenic factor I, a key regulator of steroidogenic enzyme expression, is the mouse homolog of fushitarazu-factor I. *Molecular endocrinology*, 6(8), 1249-1258.
- [22] Morohashi, K. I., Zanger, U. M., Honda, S. I., Hara, M., Waterman, M. R., & Omura, T. (1993). Activation of CYP11A and CYP11B gene promoters by the steroidogenic cell-specific transcription factor, Ad4BP. *Molecular Endocrinology*, 7(9), 1196-1204.
- [23] Hoivik, E. A., Lewis, A. E., Aumo, L., & Bakke, M. (2010). Molecular aspects of steroidogenic factor 1 (SF-1). *Molecular and cellular endocrinology*, 315(1-2), 27-39.
- [24] Lin, L. and Achermann, J.C., 2008. Steroidogenic factor-1 (SF-1, Ad4BP, NR5A1) and disorders of testis development. *Sexual Development*, 2(4-5), pp.200-209.
- [25] Arango, N. A., Lovell-Badge, R., & Behringer, R. R. (1999). Targeted mutagenesis of the endogenous mouse *Mis* gene promoter: in vivo definition of genetic pathways of vertebrate sexual development. *Cell*, 99(4), 409-419.
- [26] Sekido, R., & Lovell-Badge, R. (2008). Sex determination involves synergistic action of SRY and SF1 on a specific *Sox9* enhancer. *Nature*, 453(7197), 930-934.
- [27] Gubbay, J., Collignon, J., Koopman, P., Capel, B., Economou, A., Münsterberg, A., ...& Lovell-Badge, R. (1990). A gene mapping to the sex-determining region of the mouse Y chromosome is a member of a novel family of embryonically expressed genes. *Nature*, 346(6281), 245-250.
- [28] Koopman, P., Gubbay, J., Vivian, N., Goodfellow, P., & Lovell-Badge, R. (1991). Male development of chromosomally female mice transgenic for *Sry*. *Nature*, 351(6322), 117-121.
- [29] Albrecht, K. H., & Eicher, E. M. (2001). Evidence that *Sry* is expressed in pre-Sertoli cells and Sertoli and granulosa cells have a common precursor. *Developmental biology*, 240(1), 92-107.

- [30] Bullejos, M., & Koopman, P. (2001). Spatially dynamic expression of Sry in mouse genital ridges. *Developmental dynamics: an official publication of the American Association of Anatomists*, 221(2), 201-205.
- [31] Hacker, A., Capel, B., Goodfellow, P., & Lovell-Badge, R. (1995). Expression of Sry, the mouse sex determining gene. *Development*, 121(6), 1603-1614.
- [32] Jeske, Y. W., Bowles, J., Greenfield, A., & Koopman, P. (1995). Expression of a linear Sry transcript in the mouse genital ridge. *Nature genetics*, 10(4), 480-482.
- [33] Koopman, P., Münsterberg, A., Capel, B., Vivian, N., & Lovell-Badge, R. (1990). Expression of a candidate sex-determining gene during mouse testis differentiation. *Nature*, 348(6300), 450-452.
- [34] Bishop, C. E., Whitworth, D. J., Qin, Y., Agoulnik, A. I., Agoulnik, I. U., Harrison, W. R., ... & Overbeek, P. A. (2000). A transgenic insertion upstream of *sox9* is associated with dominant XX sex reversal in the mouse. *Nature genetics*, 26(4), 490-494.
- [35] Qin, Y., & Bishop, C. E. (2005). *Sox9* is sufficient for functional testis development producing fertile male mice in the absence of Sry. *Human molecular genetics*, 14(9), 1221-1229.
- [36] Vidal, V. P. (2001). Chaboissier MC, de Rooij DG, Schedl A. *Sox9 induces testis development in XX transgenic mice*. *Nat Genet*, 28, 216-217.
- [37] Bowles, J., Schepers, G., & Koopman, P. (2000). Phylogeny of the SOX family of developmental transcription factors based on sequence and structural indicators. *Developmental biology*, 227(2), 239-255.
- [38] Schepers, G. E., Teasdale, R. D., & Koopman, P. (2002). Twenty pairs of *sox*: extent, homology, and nomenclature of the mouse and human *sox* transcription factor gene families. *Developmental cell*, 3(2), 167-170.
- [39] Harley, V. R., Jackson, D. I., Hextall, P. J., Berkovitz, G. D., Sockanathan, S., Lovell-Badge, R., & Goodfellow, P. N. (1992). DNA binding activity of recombinant SRY from normal males and XY females. *Science*, 255(5043), 453-456.

- [40] Schmitt-Ney, M., Thiele, H., Kaltwasser, P., Bardoni, B., Cisternino, M., & Scherer, G. (1995). Two novel SRY missense mutations reducing DNA binding identified in XY females and their mosaic fathers. *American journal of human genetics*, 56(4), 862.
- [41] Li, B., Phillips, N. B., Jancso-Radek, A., Ittah, V., Singh, R., Jones, D. N., ...& Weiss, M. A. (2006). SRY-directed DNA bending and human sex reversal: reassessment of a clinical mutation uncovers a global coupling between the HMG box and its tail. *Journal of molecular biology*, 360(2), 310-328.
- [42] Harley, V. R., Layfield, S., Mitchell, C. L., Forwood, J. K., John, A. P., Briggs, L. J., ... & Jans, D. A. (2003). Defective importin β recognition and nuclear import of the sex-determining factor SRY are associated with XY sex-reversing mutations. *Proceedings of the National Academy of Sciences*, 100(12), 7045-7050.
- [43] Li, B., Zhang, W., Chan, G., Jancso-Radek, A., Liu, S., & Weiss, M. A. (2001). Human sex reversal due to impaired nuclear localization of SRY A clinical correlation. *Journal of Biological Chemistry*, 276(49), 46480-46484.
- [44] Saitou, M., Barton, S. C., & Surani, M. A. (2002). A molecular programme for the specification of germ cell fate in mice. *Nature*, 418(6895), 293-300.
- [45] Vidal, V. P., Chaboissier, M. C., de Rooij, D. G., & Schedl, A. (2001). Sox9 induces testis development in XX transgenic mice. *Nature genetics*, 28(3), 216-217.
- [46] Haber, D. A., Buckler, A. J., Glaser, T., Call, K. M., Pelletier, J., Sohn, R. L., ...& Housman, D. E. (1990). An internal deletion within an 11p13 zinc finger gene contributes to the development of Wilms' tumor. *Cell*, 61(7), 1257-1269.
- [47] Pelletier, J., Bruening, W., Li, F. P., Haber, D. A., Glaser, T., & Housman, D. E. (1991). WT1 mutations contribute to abnormal genital system development and hereditary Wilms' tumour. *Nature*, 353(6343), 431-434.
- [48] Lee, D. G., Han, D. H., Park, K. H., & Baek, M. (2011). A novel WT1 gene mutation in a patient with Wilms' tumor and 46, XY gonadal dysgenesis. *European journal of pediatrics*, 170(8), 1079-1082.
- [49] Barboux, S., Niaudet, P., & Gubler, M. C. (1997). Grünfeld JP, Jaubert F, Kuttann F, Fékété CN, Souleyreau-Therville N, Thibaud E, Fellous M, McElreavey K: Donor splice-site mutations in WT1 are responsible for Frasier syndrome. *Nat Genet*, 17, 467-470.

- [50] Hossain, A., & Saunders, G. F. (2001). The human sex-determining gene SRY is a direct target of WT1. *Journal of Biological Chemistry*, 276(20), 16817-16823.
- [51] Wilhelm, D., & Englert, C. (2002). The Wilms tumor suppressor WT1 regulates early gonad development by activation of Sf1. *Genes & development*, 16(14), 1839-1851.
- [52] Hammes, A., Guo, J. K., Lutsch, G., Leheste, J. R., Landrock, D., Ziegler, U., ...& Schedl, A. (2001). Two splice variants of the Wilms' tumor 1 gene have distinct functions during sex determination and nephron formation. *Cell*, 106(3), 319-329.
- [53] Miyamoto, N., Yoshida, M., Kuratani, S., Matsuo, I., & Aizawa, S. (1997). Defects of urogenital development in mice lacking *Emx2*. *Development*, 124(9), 1653-1664.
- [54] Kusaka, M., Katoh-Fukui, Y., Ogawa, H., Miyabayashi, K., Baba, T., Shima, Y., ...& Izuka-Kogo, A. (2010). Abnormal epithelial cell polarity and ectopic epidermal growth factor receptor (EGFR) expression induced in *Emx2* KO embryonic gonads. *Endocrinology*, 151(12), 5893-5904.
- [55] Birk, O. S., Casiano, D. E., Wassif, C. A., Cogliati, T., Zhao, L., Zhao, Y., ...& Porter, F. D. (2000). The LIM homeobox gene *Lhx9* is essential for mouse gonad formation. *Nature*, 403(6772), 909-913.
- [56] Ottolenghi, C., Moreira-Filho, C., Mendonça, B. B., Barbieri, M., Fellous, M., Berkovitz, G. D., & McElreavey, K. (2001). Absence of mutations involving the LIM homeobox domain gene *LHX9* in 46, XY gonadal agenesis and dysgenesis. *The Journal of Clinical Endocrinology & Metabolism*, 86(6), 2465-2469.
- [57] Ingham, P. W. (1998). Transducing Hedgehog: the story so far. *The EMBO journal*, 17(13), 3505-3511.
- [58] Bitgood, M. J., Shen, L., & McMahon, A. P. (1996). Sertoli cell signaling by Desert hedgehog regulates the male germline. *Current biology*, 6(3), 298-304.
- [59] Yao, H. H. C., Whoriskey, W., & Capel, B. (2002). Desert Hedgehog/Patched 1 signaling specifies fetal Leydig cell fate in testis organogenesis. *Genes & development*, 16(11), 1433-1440.
- [60] Beverdam, A., & Koopman, P. (2006). Expression profiling of purified mouse gonadal somatic cells during the critical time window of sex determination reveals novel candidate genes for human sexual dysgenesis syndromes. *Human molecular genetics*, 15(3), 417-431.

- [61] Clark, A. M., Garland, K. K., & Russell, L. D. (2000). Desert hedgehog (Dhh) gene is required in the mouse testis for formation of adult-type Leydig cells and normal development of peritubular cells and seminiferous tubules. *Biology of reproduction*, 63(6), 1825-1838.
- [62] Yao, H. H. C., Whoriskey, W., & Capel, B. (2002). Desert Hedgehog/Patched 1 signaling specifies fetal Leydig cell fate in testis organogenesis. *Genes & development*, 16(11), 1433-1440.
- [63] Colvin, J. S., Green, R. P., Schmahl, J., Capel, B., & Ornitz, D. M. (2001). Male-to-female sex reversal in mice lacking fibroblast growth factor 9. *Cell*, 104(6), 875-889.
- [64] Schmahl, J., Kim, Y., Colvin, J. S., Ornitz, D. M., & Capel, B. (2004). Fgf9 induces proliferation and nuclear localization of FGFR2 in Sertoli precursors during male sex determination. *Development*, 131(15), 3627-3636.
- [65] Kim, Y., Kobayashi, A., Sekido, R., DiNapoli, L., Brennan, J., Chaboissier, M. C., ...& Capel, B. (2006). Fgf9 and Wnt4 act as antagonistic signals to regulate mammalian sex determination. *PLoS biology*, 4(6).
- [66] Kim, Y., Bingham, N., Sekido, R., Parker, K. L., Lovell-Badge, R., & Capel, B. (2007). Fibroblast growth factor receptor 2 regulates proliferation and Sertoli differentiation during male sex determination. *Proceedings of the National Academy of Sciences*, 104(42), 16558-16563.